LIP5 Interacts with Aquaporin 2 and Facilitates Its Lysosomal Degradation

Bas W.M. van Balkom,* Michelle Boone,* Giel Hendriks,† Erik-Jan Kamsteeg,* Joris H. Robben,* H. Christiaan Stronks,* Anne van der Voorde,† Francois van Herp,§ Peter van der Sluijs,† and Peter M.T. Deen*
Departments of *Physiology and §Molecular Animal Physiology, Nijmegen Center of Molecular Sciences, and †Donders Center for Neuroscience, Radboud University Nijmegen Medical Center, Nijmegen, and †Department of Cell Biology, University Medical Center Utrecht, Utrecht, Netherlands

ABSTRACT
Vasopressin binding to the V2 receptor in renal principal cells leads to activation of protein kinase A, phosphorylation of aquaporin 2 (AQP2) at Ser256, and the translocation of AQP2 to the apical membrane, resulting in concentration of the urine. In contrast, phorbol ester–induced activation of protein kinase C pathway leads to ubiquitination of AQP2 at Lys270 and its internalization to multivesicular bodies, where it is targeted for lysosomal degradation or stored for recycling. Because little is known about the regulation of AQP2 trafficking, we used the carboxy-terminal tail of constitutively nonphosphorylated AQP2 (S256A) as a bait for interacting proteins in a yeast two-hybrid assay. We isolated lysosomal trafficking regulator–interacting protein 5 (LIP5) and found that LIP5 interacted with the proximal carboxy-terminal tail (L230-D243) of AQP2 in vitro but not with AQP3 or AQP4, which are also expressed in principal cells. Immunohistochemistry revealed that LIP5 co-localized with AQP2 in principal cells. LIP5 binding occurred independent of the state of Ser256 phosphorylation or Lys270 ubiquitination. LIP5 has been shown to facilitate degradation of the EGF receptor; here, LIP5 seemed to bind this receptor. Knockdown of LIP5 in mouse renal cells (mpkCCD) reduced the phorbol ester–induced degradation of AQP2 approximately two-fold. In summary, LIP5 binds cargo proteins and, considering the role of LIP5 in protein sorting to multivesicular bodies, plays a role in the degradation of AQP2, possibly by reducing the formation of late endosomes.


Tight regulation of the translocation of aquaporin 2 (AQP2) water channels to and from the apical membrane of renal collecting duct cells by the antidiuretic hormone arginine vasopressin (AVP) is fundamental for water homeostasis. Upon hypernatremia or hypovolemia, binding of AVP to its type 2 receptor (V2R) increases intracellular calcium and cAMP concentrations, which activate and tether protein kinase A (PKA) to AQP2-containing vesicles, resulting in phosphorylation of AQP2 and other proteins.1–7 Consequently, these vesicles dock and fuse with the apical membrane, rendering principal cells water permeable.

Regulated translocation of AQP2 to and from the apical membrane suggests the existence of proteins interacting with cytosolic segments of AQP2. Only the amino-terminal and carboxyl-terminal tails (N- and C-tails, respectively) of AQP2 extend well into the cytosol, and the C-tail of AQP2 has been shown to have an important role in its apical sorting: First, Ser256 in the AQP2 C-tail is phos-
phorylated in vivo by AVP stimulation, and studies in both cell and animal models revealed that this phosphorylation event is essential for AQP2 translocation to the plasma membrane.6–13 Second, all AQP2 mutants encoded in families with a domin-
ant inheritance of nephrogenic diabetes insipidus (NDI), a disease in which the kidney is unable to concentrate urine in response to AVP, are missorted as a result of mutations in the C-tail.12,14–19 In addition, the AQP2 C-tail is mono-ubiquiti-
nated at Lys270 upon AVP removal or PKC activation, which enhances endocytosis and degradation of AQP2.20

So far, the Rap1 GTPase-activating protein Spa1, heat-shock protein 70, and Myelin and Lymphocyte Associated Protein (MAL) are the only proteins known to bind the AQP2 C-tail, potentially playing physiologic roles in AQP2 trafficking21–23; therefore, to gain more insight in the proteins and mechanisms involved in the regulation of AQP2, we used yeast two-hybrid assays to screen a mouse kidney cDNA library for proteins inter-
acting with the C-tail of AQP2. We found the lysosomal traffick-
ing regulator (LYST) interacting protein 5 (LIP5; Swiss-Prot entry Q9CR26; corresponding gene name DRG-1) to interact specifi-
cally with AQP2. Interestingly, LIP5 is reported to function in

Site and Specificity of the AQP2–LIP5 Interaction
To allow further characterization of the role of LIP5 in AQP2 binding and regulation, we generated antibodies directed against full-length LIP5. Immunohistochemical/cytochemical and immunoblot data (Supplemental Figure 2) and the immu-
oblot data from the LIP5 shRNA experiment reveal the spec-
ificity of our LIP5 antibodies.

RESULTS

Screening for AQP2-Interacting Proteins
To isolate proteins involved in AQP2 regulation, we trans-
fected yeast cells expressing (for clarity, expression refers to
protein expression unless indicated otherwise) a fusion protein of LexA and the C-tail of AQP2-S256A with a mouse kidney cDNA library, grown under selective conditions and screened for β-galactosidase activity. Approximately 3.85 × 10^6 colo-

LIP5 Interaction with the EGF Receptor
Our data reveal AQP2 as the first cargo protein identified to
interact with the AQP2 tail, but, again, not with GST-AQP4 or GST (Figure 1B).

LIP5 Expression in the Kidney
For LIP5 to have a role in the regulation of AQP2 in vivo, it
needs to be present in renal principal cells. We performed LIP5
in situ hybridization and AQP2 immunohistochemistry on alternating mouse kidney sections. Microscopic analysis revealed that renal principal cells (positive for AQP2; Figure 2A, 2 and 4) indeed also contain LIP5 mRNA (Figure 2A, 1 and 3). Besides these cells, LIP5 mRNA was detected in epithelial cells of other renal tubules and collecting duct cells. A sense probe of LIP5 cRNA, which was taken along as a negative control, revealed no staining (Figure 2A, 5).

Next, AQP2 and LIP5 co-localization was determined. Immunohistochemistry for AQP2 and LIP5 on mouse kidney sections and confocal laser scanning microscopy revealed that AQP2 co-localizes with LIP5 in the apical region of renal principal cells (Figure 2B). Interestingly, in AQP2-negative tubules (arrows in Figure 2B) and intercalating cells (asterisks in Figure 2B), which are the AQP2-negative cells of collecting ducts, LIP5 showed a punctuate staining, suggesting a vesicular localization of LIP5 in these cells.

Figure 1. Interaction of LIP5 with AQP2, AQP3, AQP4, and EGFR. (A) Yeast cells expressing LexA fused to the C-tail of wt-AQP2, AQP2-S256A, AQP2-S256D, AQP2-E258K, AQP3, or AQP4 proteins, all in combination with GAL4-LIP5 fusion proteins, were analyzed for interaction using β-galactosidase activity assays. Yeast cells expressing the C-tails of wt-AQP2, AQP2-S256A, AQP2-S256D, and AQP2-E258K but not those of AQP3 or AQP4 show blue staining of the colonies and thus interaction with LIP5. Sequential stop mutants of the C-tail of AQP2 revealed that only upon deletion of region L230-D243, binding of the AQP2 C-tail with LIP5 is lost. Control yeast cells transfected with bait and empty prey constructs did not show any β-galactosidase activity. (B and C) GST or GST fused to the C-tails of AQP2 (GST-AQP2) or AQP4 (GST-AQP4) were incubated with in vitro translated LIP5 (B) or dog kidney cytosol (C) and subjected to GST pulldown assays. LIP5 was visualized by autoradiography (B) or immunoblotting (C). (D) For determination of whether LIP5 also interacts with the EGFR, Xpress-tagged LIP5 was expressed alone or together with the EGFR (in duplicate) in HEK293 cells, lysed, and subjected to EGFR immunoprecipitation. Subsequent immunoblotting of the precipitates using mouse anti-Xpress-tag antibodies revealed that Xpress-tagged LIP5 co-precipitated with the EGFR but was not precipitated when expressed alone (top). EGFR immunoblotting of the lysates revealed specific signals for the EGFR in lanes of cells transfected with EGFR constructs (bottom). LIP5 immunoblotting showed Xpress-tagged (exo) and endogenous (endo) LIP5 in cells transfected with LIP5 constructs (third panel).

Effect of Physiologic Modification in the AQP2 C-Tail on Its Interaction with LIP5

In vivo, redistribution of AQP2 from intracellular vesicles to the apical membrane coincides with phosphorylation of AQP2 at Ser256.10,11 Subsequent studies in MDCK cells showed that phosphorylation at Ser256 is essential and sufficient for apical membrane localization of AQP2, because AQP2-S256A, which mimics nonphosphorylated AQP2, is localized in intracellular vesicles, whereas AQP2-S256D, mimicking phosphorylated AQP2, is localized in the apical membrane.8 Also, AQP2 mutants in dominant NDI have missense or frame-shift mutations in the C-tail, one of which is AQP2-E258K.15,16 For determination of whether LIP5 binding to the AQP2 C-tail depends on the AQP2 phosphorylation state and whether binding is lost with AQP2 mutants in dominant NDI, the C-tails of AQP2 were expressed with LIP5 in yeast and tested for interaction. Co-immunoprecipitation assays and subsequent immunoblotting revealed that LIP5 binds both AQP2-S256A and AQP2-S256D, and AQP2-E258K were expressed with LIP5 in yeast and tested for interaction. β-Galactosidase assays revealed a positive signal for the S256A, S256D, and E258K C-tails (Figure 1A), suggesting that LIP5 binding is independent of the AQP2 phosphorylation state and is maintained in the AQP2-E258K mutant. Using HEK293 cells, transiently-expressing AQP2-S256A or AQP2-S256D, co-immunoprecipitation assays confirmed that interaction of LIP5 with AQP2 is independent of AQP2 phosphorylation state (Figure 3A).

Besides phosphorylation, AQP2 is subject to mono-ubiquitination at Lys270, which enhances endocytosis and degradation of AQP2.20 To determine whether LIP5 interaction with AQP2 depends on the ubiquitination state of AQP2, wt-AQP2, AQP2 that cannot be ubiquitinated (AQP2-K270R), or constitutively ubiquitinated AQP2 (AQP2-Ub) were expressed in HEK293 cells and tested for interaction. Co-immunoprecipitation assays and subsequent immunoblotting revealed that LIP5 binds both AQP2-K270R and AQP2-Ub, indicating that LIP5 binding to AQP2 is independent of AQP2 ubiquitination (Figure 3B).

Role of LIP5 in AQP2 Abundance

LIP5 is implicated in sorting of proteins to the internal vesicles of MVBs after endocytosis, from where proteins are targeted
To investigate the effect of TPA on the half-life of AQP2, we incubated vasopressin-stimulated mpkCCD cells for various periods with or without TPA in the continuous presence of cycloheximide and immunoblotted for AQP2 (Figure 4C). After 60 min of TPA treatment, AQP2 levels were significantly ($P < 0.05$) decreased for TPA-treated versus control cells. Calculations revealed that TPA reduced the half-life of AQP2 from 234.5 ± 32.7 to 66.2 ± 14.3 min.

To investigate whether LIP5 plays a role in AQP2 degradation, we made lentiviruses driving the expression of two different murine LIP5 (LIP5–1/2) shRNAs or a random sequence (mock). For recognition of infected cells, the viral DNA also encoded cytomegalovirus (CMV) promoter–driven GFP. Testing the viruses in untransfected, dDAVP-stimulated mpkCCD cells revealed that most cells had lost the shRNA constructs at the time of appropriate AQP2 abundance (at 4 d of dDAVP treatment after 4 d of cell polarization); therefore, mpkCCD cell lines stably expressing AQP2 were generated. To perform experiments with physiologically relevant AQP2 protein levels, we selected clones that express AQP2 at a similar or lower level than endogenous AQP2 after stimulation with 1 nM dDAVP by immunoblotting (data not shown). After infection of mpkCCD-AQP2 cells with shRNA viruses, immunocytochemistry, subsequent immunoblot analysis for LIP5 revealed that LIP5 shRNAs but not mock shRNA yielded significantly reduced LIP5 protein levels (90 and 70% for LIP5-1 and LIP5-2, respectively; Figure 5A), whereas cells infected with a virus encoding the mock shRNA (Figure 5A) showed no difference in LIP5 protein abundance compared with uninfected cells. This indicated that LIP5-specific shRNAs but not viral infection itself affects LIP5 protein abundance. Unfortunately, we did not succeed in co-staining for LIP5 and AQP2 in the GFP background.

To determine the effect of LIP5 knockdown on AQP2 degradation, we treated cells with cycloheximide in the absence or presence of TPA for 2 h and lysed them. Consistent with immunocytochemistry, subsequent immunoblot analysis for LIP5 revealed that LIP5 shRNAs but not mock shRNA yielded significantly reduced LIP5 abundance in cells infected with the LIP5-specific shRNAs (LIP5–1, LIP5–2; Figure 5A), whereas cells infected with a virus encoding the mock shRNA (Figure 5A) showed no difference in LIP5 protein abundance compared with uninfected cells. This indicated that LIP5-specific shRNAs but not viral infection itself affects LIP5 protein abundance.
Figure 3. LIP5 interaction with physiologically modified AQP2. (A) For determination of whether LIP5 binding to AQP2 depends on the phosphorylation state of AQP2, LIP5 was expressed together with wt-AQP2, AQP2-S256D, and AQP2-S256A in HEK293 cells; lysed; and subjected to LIP5 immunoprecipitation. Subsequent immunoblotting of the precipitates demonstrated that wt-AQP2, AQP2-S256D, and AQP2-S256A co-precipitated with LIP5 (top). Immunoblotting of the lysates revealed similar signals for AQP2 in lanes of cells transfected with AQP2 protein constructs (middle) and for endogenous (endo) and exogenous (exo) LIP5 in cells transfected with LIP5 constructs (bottom). (B) For determination of whether LIP5 binding to AQP2 depends on the ubiquitination state of AQP2, LIP5 was expressed together with wt-AQP2, AQP2-K270R, and AQP2-Ub in HEK293 cells. Subsequent cell lysis and LIP5 immunoprecipitation followed by immunoblotting showed that wt-AQP2, AQP2-K270R, and AQP2-Ub co-precipitated with LIP5 (top). Immunoblotting of the lysates revealed equal signals for AQP2 in lanes of cells transfected with wt-AQP2, AQP2-K270R, and AQP2-Ub constructs (middle) and for endogenous (endo) and exogenous (exo) LIP5 in cells transfected with LIP5 constructs (bottom).

DISCUSSION

LIP5 Specifically Interacts with the Proximal Region of the AQP2 C-Tail Independent of AQP2 Phosphorylation or Mono-ubiquitination

We identified LIP5 to interact with the C-tail of AQP2-S256A in yeast two-hybrid assays. Whereas AQP2 is coexpressed with AQP3 and AQP4 in renal principal cells and all have a similar gross architecture, yeast two-hybrid and GST pulldown analyses revealed that LIP5 interacts directly and specifically with the AQP2 C-tail only, not with the C-tails of AQP3 or AQP4 (Figure 1, A through C). Further analysis of the AQP2 C-tail revealed that LIP5 binds region L230 to D243 of AQP2 (Figure 1A). Consistent with binding to this region of AQP2, the interaction of LIP5 to AQP2 is independent of AQP2 phosphorylation (at Ser256) and mono-ubiquitination (at K270) and is also not affected by an AQP2 mutation causing NDI, because the sites of these modifications are outside the LIP5 binding region (Figure 3). The precise mode of interaction and amino acids involved remains to be established.

LIP5 Interacts with Cargo Proteins

The MVB machinery is composed of three complexes—ESCRT-I, -II, and -III—which are sequentially recruited to sites of MVB sorting and vesicle formation. Subsequent release of ESCRT-III from the membrane requires Vps4 ATPase activity and allows the ESCRT machinery to recycle through multiple rounds of luminal vesicle formation. So far, LIP5 has been shown to interact with Vps4 and several ESCRT-III components, including CHMP5, CHMP1B, CHMP2A, and CHMP3.24,29–33 Our data reveal for the first time that LIP5 also interacts with cargo proteins, because AQP2 as well as the EGFR specifically co-precipitated with LIP5 (Figure 1). It will be interesting to unravel to which LIP5 segment these two proteins bind and whether these two proteins compete with CHMPs for binding to Vps4.

LIP5 Is Involved in the Lysosomal Degradation Pathway of AQP2

Membrane proteins targeted for lysosomal degradation are sorted into vesicles that bud into MVBs. These MVBs can serve as long-term storage compartments, fuse with lysosomes to deliver the internal vesicles and their contents for degradation, or fuse with the plasma membrane to release the vesicles as extracellular exosomes. MVB vesicle formation and protein sorting require a set of class-E vacuolar protein sorting (VPS) proteins.34–36 Most class-E proteins function as components of one of the three ESCRT complexes, which are sequentially recruited to sites of MVB sorting and vesicle formation. The ESCRT-III proteins are the last to assemble, forming a membrane-associated lattice that functions in the final stages of this process. Via direct protein–protein interactions, Vps4 is recruited to MVBs, which induces the release of the ESCRT-III complex from the membrane.34,36

Emerging evidence points toward a role for LIP5 dimers in the disassembly of the ESCRT-III complex by stimulating Vps4 ATPase activity, which occurs directly through interaction between the conserved C-terminal VSL domain of LIP5 and Vps4.31–33 Consistently, deletion of Vta1 (the yeast LIP5 orthologue) in yeast results in altered vacuolar morphology,29 and knockdown of LIP5 in mammalian cells facilitates downregulation of EGFR.24

Several data suggest a similar role for LIP5 in AQP2 regulation in principal cells: First, LIP5 is coexpressed with AQP2 in renal principal cells (Figure 2) and LIP5 has been identified in AQP2-containing exosomes isolated from urine (Myo12 protein).37 Second, renal LIP5 interacts specifically with the C-terminal tail of AQP2 (Figure 1). Third, LIP5 facilitates the lysosomal degradation of AQP2: Whereas induced LIP5 overexpression in MDCK-AQP2 cells did not affect AQP2 abundance (Supplemental Figure 3) or translocation to the apical membrane, knockdown of LIP5 abundance resulted in a 1.5- to 2.5-fold increase in the AQP2 half-life after TPA treatment (Figure 5, B and C), a condition known to induce AQP2 internalization and degradation8 (Figure 4). Considering our find-
ing that AQP2 levels seemed somewhat increased in LIP5 shRNA versus mock cells without TPA treatment (Figure 5, B and C), the half-life of AQP2 in LIP5 knockdown cells without TPA treatment may be higher than that of AQP2 in dDAVP-stimulated mpkCCD cells (Figure 4C); therefore, the estimated increase in AQP2 half-life with LIP5 knockdown after TPA treatment should be taken as a rough estimate.

Under the tested conditions, the effect of LIP5 knockdown is clear upon treatment with TPA but not without TPA treatment, which can be explained as follows: At any moment in time, AQP2 resides at several locations in the cell (endoplasmic reticulum, Golgi, storage vesicles, plasma membrane, recycling vesicles, MVBs, and lysosomes), and the steady-state localization of AQP2 is a balance regulated by extracellular and intracellular signals, such as the presence of AVP, and cAMP-dependent phosphorylation or activation of the PKC pathway. TPA-induced degradation of AQP2 in mpkCCD cells mainly occurs via lysosomes (Figure 4B) and, as illustrated by the large half-life of AQP2 of nearly 4 h (Figure 4C), only a small fraction of AQP2 is targeted for lysosomal degradation in the tested 2-h period under non-TPA conditions. With the TPA-induced internalization and lysosomal degradation of AQP2, which decreases the AQP2 half-life to approximately 70 min, more AQP2 will thus pass LIP5 on its way to MVBs/lysosomes for the 2 h measured, and, therefore, the effect of (the absence of) LIP5 on AQP2 is larger and better detectable under these conditions. The partial inhibition of AQP2 degradation in our LIP5 shRNA experiments may suggest that, like its yeast orthologue Vta1,33 LIP5 has only a modulatory role in the lysosomal degradation of AQP2; however, this cannot be deduced from our experiments, because as a result of incomplete infection and/or shRNA knockdown, our shRNA-expressing cells were not devoid of LIP5 (Figure 5B).

On the basis of these data, we propose the following model for the role for LIP5 in the regulation of AQP2 in principal cells: After stimulation with AVP and translocation of AQP2 to the apical membrane, renal water reabsorption will occur, which may be increased by an extended presence of AQP2 in the plasma membrane through an interaction with MAL.22 With removal of AVP or activation of the PKC pathway, which is thought to follow binding of hormones such as endothelin, PGE2, and ATP, AQP2 is ubiquitinated at Lys270, which signals its endocytosis.20 After recruitment to clathrin-coated vesicles, AQP2 is then endocytosed from the apical membrane involving direct interaction with heat-shock protein 70.23 Via early endosomes, AQP2 is then sorted via ESCRT proteins and interaction with LIP5 to inner vesicles of MVBs, from where it can be targeted to lysosomes for degradation.
or released as exosomes from the cells into urine (Figure 6).37 Unfortunately, we were not able to co-immunoprecipitate AQP2 with LIP5 from mouse kidney homogenates, which may be due to a reduced abundance of LIP5 or low level of AQP2–LIP5 interaction in the kidney.

The binding of LIP5 to the proximal region of the AQP2 C-tail and its independence of the ubiquitination and phosphorylation status of AQP2 (Figure 3) is in line with the finding that TPA-induced internalization of AQP2 constitutively phosphorylated at S256 still leads to its degradation38 and that AQP2 degradation is well detectable only at 2 h after TPA treatment, at which point AQP2 is not detectably ubiquitinated anymore.20 Recently, AQP2 was found to be phosphorylated at other sites in its C-terminus.39 It remains to be established whether phosphorylation of these sites influences LIP5 binding to AQP2.

Interestingly, LIP5 has also been reported to interact with the lysosomal trafficking regulator LYST,40 which is a cytosolic protein of 425 kD with a putative function in lysosome-related organelles.41 Yeast two-hybrid screens with LYST identified several putative partners, some of which have been shown to co-localize (HRS)42 or be involved (calmodulin)1 in the regulation of AQP2. If LYST is expressed in collecting duct cells, then LYST may be a scaffolding protein involved in the degradation of AQP2.

Moreover, calmodulin has been shown to interact with proximal C-terminal tail of AQP0,43 the region bound by LIP5 in AQP2. Although these regions are similar between AQP0 and AQP2, we could not detect binding between heterologously expressed calmodulin and the AQP2 C-tail in GST pulldown experiments (data not shown).

In summary, we have identified that LIP5 is co-expressed with AQP2 in renal principal cells, interacts with the proximal C-tail of AQP2, and facilitates its lysosomal degradation. Our data thus indicate that LIP5 plays an important role in the MVB/lysosomal targeting of AQP2, after which it will be degraded or expelled from the cells into urine. It will be interesting to see how LIP5 structurally couples to AQP2, whether the degradation of other cargo proteins is facilitated by direct interaction with LIP5, and whether LIP5 also affects AQP2 stability in vivo.

CONCISE METHODS

Yeast Two-Hybrid
cDNAs encoding wild-type and mutant AQP2 C-termini starting at amino acid Phe224 were generated by standard PCR reactions using AQP2-C-EcoRI (5’-GATGAAAATTTCGCGCCAGCCAAGAGCCT-3’) as a forward primer and AQP2-C-Hi as a reverse primer. As templates, pCB6-wtAQP2, pCB6-AQP2-S256A, and pCB6-AQP2-S256D were used. cDNAs encoding truncated AQP2 C-tail proteins were generated similarly using AQP2-C-EcoRI as a forward primer and AQP2-R252X (5’-GATGCAATTCGTCACTGGATCGCGT3’), AQP2-D243X (5’-GATGCAATTCGTCACTGGATCGCGT3’), and AQP2-R252X (5’-GATGCAATTCGTCACTGGATCGCGT3’) as reverse primers.
Cytosol of dog kidneys was prepared by crushing the tissue in a Promix; Amersham Biosciences, Uppsala, Sweden) and incubated in vitro transcription/translation using the Riboprobe-T7 system (Promega, Leiden, Netherlands) in the presence of 35S-methionine/cysteine (Redivue Pharmaceuticals, Palo Alto, CA) against a bait of the AQP2-S256A C-terminal tail. For library of adult mouse whole kidneys (MATCHMAKER, Clontech, Palo Alto, CA) screening, four independent colonies was induced by IPTG and isolated with prewashed Glutathione-Sepharose 4B beads. LIP5 was produced by in vitro transcription/translation using the Riboprobe-T7 system (Promega, Leiden, Netherlands) in the presence of 35S-methionine/cysteine (Redivue Pharmaceuticals, Uppsala, Sweden) and incubated with the GST fusion protein beads, rotating overnight at 4°C. Bound LIP5 was eluted by boiling the samples in Laemmli sample buffer and analyzed by SDS-PAGE. Pulldown LIP5 was visualized using a STORM phosphor imager (Amersham Biosciences, Freiburg, Germany).

GST, GST-AQP2, and GST-AQP4 production in BL21-DE3 bacteria was induced by IPTG and isolated with prewashed Glutathione-Sepharose 4B beads. LIP5 was produced by in vitro transcription/translation using the Riboprobe-T7 system (Promega, Leiden, Netherlands) in the presence of 35S-methionine/cysteine (Redivue Pharmaceuticals, Uppsala, Sweden) and incubated with the GST fusion protein beads, rotating overnight at 4°C. Bound LIP5 was eluted by boiling the samples in Laemmli sample buffer and analyzed by SDS-PAGE. Pulldown LIP5 was visualized using a STORM phosphor imager (Amersham Biosciences, Uppsala, Sweden).

The GST-LIP5 expression construct, pACT2 clone 14b (identified from library screening) was cut with EcoRI and Xhol and cloned into corresponding sites of pGEX4T-1 (Amersham Biosciences, Freiburg, Germany). For GST-AQP4 expression, we used a pGEX1 vector containing the C-tail of AQP4. To make a GST-LIP5 expression construct, pACT2 clone 14b (identified from library screening) was cut with EcoRI and Xhol, and the cDNA fragment encoding full-length LIP5 was isolated and cloned into the corresponding sites of pGEX6.1 (Amersham Biosciences, Freiburg, Germany).

GST, GST-AQP2, and GST-AQP4 production in BL21-DE3 bacteria was induced by IPTG and isolated with prewashed Glutathione-Sepharose 4B beads. LIP5 was produced by in vitro transcription/translation using the Riboprobe-T7 system (Promega, Leiden, Netherlands) in the presence of 35S-methionine/cysteine (Redivue Pharmaceuticals, Uppsala, Sweden) and incubated with the GST fusion protein beads, rotating overnight at 4°C. Bound LIP5 was eluted by boiling the samples in Laemmli sample buffer and analyzed by SDS-PAGE. Pulldown LIP5 was visualized using a STORM phosphor imager (Amersham Biosciences, Uppsala, Sweden).
leupeptin, and 5 µg/ml pepstatin using a potter (PorrterS, Braun Bio-
tech, Melssun, Germany). Cytosol was cleared by centrifugation at
33,000 rpm in a Ti45 rotor in a Beckman ultracentrifuge for 1 h at 4°C.
The supernatant was preincubated for 1 h with glutathione-Sepharose
4B beads containing GST and subsequently incubated with GST,
GST-AQP2, or GST-AQP4 glutathione-Sepharose 4B beads over-
night at 4°C. After binding, the beads were washed with 25 mM Tris
(pH 7.5), 0.5 mM EDTA, and 1% Triton-X-100, and bound proteins
were eluted by shaking in 1.5 M NaCl, 25 mM Tris (pH 7.5), and 0.5
mM EDTA at room temperature for 1 h. Samples were desalted using
Biospin columns (Biorad, Hercules, CA). Isolated proteins were ana-
alyzed by SDS-PAGE and immunoblotting.
Empty or transfected COS cells were lysed in 300 µl of lysis buffer
with 1% NP40. For LIP5 depletion, lysates were incubated for 2 h with
4 µl of LIP5 antiserum and 40 µl of protein A-agarose beads (Kem-
En-Tec A/S, Copenhagen, Denmark). Lysates were then incubated with
GST-AQP2 glutathione-Sepharose 4B beads and further pro-
cessed as described already.

Expression in Eukaryotic Cells
For expression of LIP5 in eukaryotic cells, an EcoRI-Xhol fragment
encoding full-length LIP5 was isolated from the pACT2 clone 14b, iden-
tified from library screening, and ligated into the corresponding
sites of pCDNA3. Expression constructs encoding wild-type AQP2 or
the S256A, S256D, K270R, and AQP2-UB mutants were as described
previously.2,20 pCDNA3-EGFR was provided by Prof. Y. Yarden (De-
partment of Biologic Regulation, Weizmann Institute of Science, Re-
hovot, Israel).51 HEK293 cells were maintained in DMEM (Biovit-
taker, Verviers, Belgium) supplemented with 10% FCS. For transient
transfections, HEK293 cells were seeded at 1.2 × 10⁵ cells/cm² in
six-well plates. Four hours after seeding, cells were transfected using
polyethylenimine (Polyscience, Eppelheim, Germany). A total of 3.5
µg of circular DNA and 14 µl of polyethylenimine (1 µg/µl) were
added to 80 µl of OptiMEM medium (Life Technologies Europe,
Breda, Netherlands), vortexed, and incubated at room temperature
for 20 min. Subsequently, the mixture was added to the cells and
incubated overnight. Three days after transfection, cells were lysed
in IP lysis buffer (1% Triton X-100, 150 mM NaCl, and 25 mM HEPES
[pH 7.4]) and incubated with rabbit anti-LIP5 antibodies, mouse
anti-Xpress, or mouse anti-EGFR 528 (Santa Cruz Biotechnology,
Copenhagen, Denmark) immobilized on protein A-agarose beads (Kem-
En-Tec A/S) overnight at 4°C. Immunoprecipitated proteins were ana-
alyzed by immunoblotting.

In Situ Hybridization
LIP5 cDNA templates with a T7 RNA polymerase promoter sequence
(underlined in primer sequences) at the 5’ or 3’ end were used to generate
LIP5 cRNA probes. These templates were made by a standard PCR
reaction, using 5’-TATACACTCTACTATAGGAGAGGCCCTTCT-
GCGGCGCTG-3’ or 5’-CGTACGGTCAGCTGCTACTGTTGCGT-
ATACA-3’ (sense) together with 5’-GGGCCCCCTGCAGCGCGTGC-3’
or 5’-TATAGGACACTCATAAGGAGAGCTCGATCTGCTT-
CATGTGCTGATACA-3’ (antisense) primers, respectively. 11-UTP
dioxigenin (DIG)-labeled probes of these templates were made in vitro
using T7 RNA polymerase (Roche Diagnostics, Alamere, Netherlands).
For preparation of mouse kidney sections, mice were transcardially
perfused with ice-cold 0.6% NaCl solution for 5 min, followed by
Bouin’s fixative for 15 min. After dissection, kidneys were postfixed in
Bouin’s fixative for 24 h. Then, kidneys were dehydrated in a graded
series of ethanol and xylene and embedded in paraffin. Seven-mi-
crometer sections were mounted on poly-l-lysine–coated slides and
left for 16 h at 37°C before they were rehydrated in a graded series of
ethanol. Tissue penetration was enhanced by incubation in 0.1% pep-
sin in 0.2 M HCl for 15 min at 37°C, followed by fixation in 4% parafo-
raldehyde in PBS for 5 min and incubation in 1% hydrox-
ylaminiumchloride for 10 min. After this, sections were dehydrated in
ethanol and air dried. Hybridization took place for 1 h at 55°C in
hybridization buffer (10% sodium dextran sulfate, 50% formamidine,
4× salt and sodium citrate (SSC), 1× Denhardt’s and 200 µg/ml yeast
tRNA; 1× SSC = 0.15 M NaCl and 0.015 M sodium citrate) with 500
ng/ml DIG-labeled probe. After stringency washes in 2× SSC, 1× SSC,
0.5× SSC, and 0.1× SSC for 30 min at 37°C, sections were rinsed
for 10 min in Tris-buffered saline (TBS), blocked in blocking solution
(1% BSA and 2% normal goat serum in TBS) for 30 min, and incu-
bated with alkaline phosphatase (AP)-conjugated sheep anti-DIG Fab
fragments (1:500; Roche Diagnostics) in blocking solution for 16 h at
4°C. After three washes of 10 min in TBS and one wash of 5 min in AP
buffer (100 mM Tris and 100 mM NaCl [pH 9.5]), sections were
stained in 350 µg/ml 4-nitro blue tetrazolium chloride and 175 µg/ml
5-bromo-4-chloro-3-indolyl-phosphate (Roche Diagnostics) in AP
buffer until color development was sufficient. Then, sections were
washed twice with distilled H₂O and mounted in Mowiol. Alternating
sections were used for AQP2 immunohistochemistry. After mounting
of fixed sections on poly-l-lysine–coated slides and rehydration in
graded series of ethanol, sections were washed with TBS, incubated
with 1% H₂O₂ in TBS for 15 min to quench endogenous peroxidase
activity, rinsed with TBS, incubated with 1:100 diluted rabbit anti-
AQP2 antibodies in blocking buffer, washed three times with TBS,
and incubated for 1 h in 1:100 diluted donkey anti-rabbit antibodies
coupled to horseradish peroxidase (Jackson ImmunoResearch Lab-
ratories, West Grove, PA). After washing three times in TBS, sections
were incubated in diaminobenzidine until color development was
sufficient. Then, sections were dehydrated in graded series of alcohol
and ethylene and mounted in Entellan (Merck, Darmstadt, Ger-
many). A sense LIP5 cRNA probe was taken along as a negative con-
control for the specificity of the LIP5 mRNA hybridization.

mpkCCD Cells and shRNA Assays
MpkCCD cells (clone 14²⁷) were grown in modified defined medium
(DMEM:Ham’s F12: 1:1 vol/vol; 60 mM sodium selenate, 5 µg/ml
transferrin, 2 mM glutamine, 50 nM dexamethasone, 1 nM triodo-
thyronine, 10 ng/ml EGF, 5 µg/ml insulin, 20 mM t-glucose, 2% FCS,
and 20 mM HEPES [pH 7.4]) at 37°C in an air atmosphere of 5% CO₂.
The medium was replaced every 2 d. Exponentially growing cells (at
approximately 70% confluence) were trypsinized and seeded at a den-
sity of 1.5 × 10⁵ cells/cm² on semipermeable filters (Transwell,
0.4-µm pore size; Corning Costar, Cambridge, MA) of 1.13 cm². The
cells remained in culture for a total period of 8 d before being ana-
lyzed. The cells were treated for the last 96 h with 1 nM dDAVP to only
the basolateral side to induce AQP2 expression maximally. TPA (0.1
μM), cycloheximide (50 μM), MG132 (20 μM), and chloroquine (100 μM) were administered to both the apical and the basolateral compartments for the indicated time periods.

To make pLV-CMV-GFP-shRNA-LIP5 constructs for the LIP5 shRNA assays, we first created the pTER-LIP5 construct. Phosphorylated LIP5-specific oligonucleotides (10 μg) were annealed in 100 μl of 25 mM KCl by incubating the mixture at 95°C for 2 min and slowly cooling to room temperature. A total of 1 μl of the mixture was ligated into the BglII- and HindIII-digested pTER vector. The following oligonucleotides were used: 5’-GATCCCGGACAGTAGACTCTGTTGTTCAAGAGACC-3’, 5’-CTTGAACCACACAGGAGTCATTGTCGCGG-3’, 5’-ACACAGGACTTGTCTTTTTGAAA-3’, and 5’-AGCTTTTCTAAAAAGACATCCTGTTGCTTCT-3’ for pTER-LIP5-1; 5’-GATCCCGGACAGTAGACTCTGTTGCTTCTGCAGCATGTTGCTTCT-3’ for pTER-LIP5-2. To generate the lentiviral LIP5 shRNA constructs, we inserted the PstI fragments from the pTER-LIP5 constructs containing the H1 promoter and LIP5 shRNA sequences into the corresponding site of pLV-CMV-GFP.53

MpkCCD cells were stably transfected with pCB6-AQP2 using the calcium-phosphate precipitation technique as described previously.54 Third-generation lentiviruses were produced by co-transfection of the packaging vectors pRSV-Rev, pMDL g/p RRE, and pMD2G (Tronolab, Lausanne, Switzerland) and the transfer vector pLV-CMV-GFP-shRNA-LIP5 into human embryonic kidney 293T cells as supported by grants from the Dutch Organization of Scientific Research (NWO). This research was supported by grants from the European Union (RTN aquaglyceroporins; 035995-2), kidney foundation (C03.2060), UMC St Radboud (2004-55) and NWO (865.07.002) to P.M.T.D.

ACKNOWLEDGMENTS

P.M.T.D. is a recipient of a VICI grant (865.07.002) of the Netherlands Organization for Scientific Research (NWO). This research was supported by grants from the Dutch Organization of Scientific Research (NWO-MW 902-18-292) to P.M.T.D. and P.v.d.S., and from the European Union (RTN aquaglyceroporins; 035995-2), kidney foundation (C03.2060), UMC St Radboud (2004-55) and NWO (865.07.002) to P.M.T.D.

We thank Ronnie Wismans and Tony Coenen for superb technical support and Dr. S. Gisler (Department Physiology, University of Zurich, Zurich, Switzerland) for sending us material and sharing his technical expertise on our yeast two-hybrid analyses.

DISCLOSURES

None.

REFERENCES


34. Babst M: A protein’s final ESCRT. Traffic 6: 2–9, 2005


Supplemental information for this article is available online at [http://www.jasn.org/](http://www.jasn.org/).